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Synthesis and anti-inflammatory activity of *N*-phthalimidomethyl 2,3-dideoxy- and 2,3-unsaturated glycosides

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Abstract—3,4,6-Tri-*O*-acetyl-D-galactal, 3,4,6-tri-*O*-acetyl-D-glucal and 3,6,2′,3′,4′6′-hexa-*O*-acetyl-D-lactal were reacted with *N*-hydroxymethylphthalimide and boron trifluoride etherate to produce the corresponding phthalimidomethyl unsaturated glycosides via Ferrier rearrangement. When the galactal derivative was used, a non-Ferrier rearrangement product was also isolated as a minor product under classical Ferrier conditions. Phthalimidomethyl deoxy glycosides were readily prepared by hydrogenation of the unsaturated glycosides. Following deacetylation, the anti-inflammatory activities of these compounds were tested on mice and three were found to possess potent activity compared to hydrocortisone sodium succinate (HSS).

mice.

Keywords: Phthalimidomethyl sugar derivatives; Glycal; Deoxy-sugar; Anti-inflammation

1. Introduction

The phthalimido group is a common protecting group for amines and is widely used in carbohydrate chemistry. As a known pharmacophore, the phthalimido moiety also has a wide range of activities such as anti-tumor, ¹ anti-convulsant,² and anti-inflammatory activity.³ Systematic investigation of phthalimide analogues revealed that derivatives bearing a spherical alkyl group, such as an adamantyl group or a carboranyl group, possess potent bi-directional TNF- α (tumor necrosis factor- α) production-regulating activity.4 Recently, the synthesis of *N*-phthalimidomethyl tetra-O-acyl-α-D-mamopyranoside derivatives and their hypolipidemic activity was reported.⁵ However, the bioactivities of N-phthalimidomethyl glycosides have not been fully studied. As the phthalimido moiety is insoluble in water, its conjugation to sugars bearing free hydroxyl groups may enhance their water-solubility, thus making the whole

structure more 'drug-like'. Furthermore, the carbohydrate moiety could play an important role in directing

the pharmacophore to a specific site in vivo. With these

ideas in mind, a series of phthalimidomethyl 2,3-dideoxy

sugars and 2,3-unsaturated sugars were designed and

synthesized to study the activity of phthalimide deriva-

tives. Their anti-inflammatory activities were tested on

3,4,6-tri-*O*-acetyl-D-glucal **2b**, or 3,6,2',3',4'6'-hexa-*O*-

^{2.} Results and discussionGlycals are versatile precursors in oligosaccharide synthesis. In our experiments, *N*-phthalimidomethyl 2,3-unsaturated sugars **3a**, **3b**, and **9** were synthesized through nucleophilic addition of *N*-hydroxymethyl phthalidomide **1** to 3,4,6-tri-*O*-acetyl-D-galactal **2a**,

acetyl-D-lactal $\bf 8$ in the presence of boron trifluoride etherate, resulting in a Ferrier rearrangement (Scheme 1).⁷ Although it is known that the 2,3-unsaturated sugars formed by this reaction are usually obtained as an anomeric mixtures, only the α -anomer was obtained in our work. The anomeric configurations of compound $\bf 3a$, $\bf 3b$,

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Scheme 1. Synthesis of N-phthalimidomethyl 2,3-dideoxy-glycosides and 2,3-unsaturated glycosides.

and **9** were confirmed by NMR spectroscopy. Srivastava et al. had also obtained exclusively the α -anomer in the similar reaction starting from 3,4,6-tri-O-acetyl-D-glucal.⁵

The Ferrier rearrangement product **3a** was isolated as the major product contaminated with a byproduct, which was rather difficult to be separate from the mixture. By very careful column chromatography, this byproduct was obtained pure in 10–30% yields from different batches. Based on its NMR and MS data, the structure of this product was assigned as phthalimidomethyl 3,4,6-tri-*O*-acetyl-2-deoxy-α-D-*lyxo*-hexopyranoside, **4a**. Lin et al. hvae also reported similar results. Reports by Ferrier and Wallenfels concluded that the rearrangement product could only be produced in the absence of protic acids whereas in the presence of protic acids addition to the double bond would occur to yield

2-deoxy sugar derivatives. In our case, the 2-deoxy product only occurred with 3,4,6-tri-O-acetyl-D-galactal. We suggest a proton first adds to the double bond as a competitive side reaction of Ferrier rearrangement to give the 2-deoxy oxacarbenium ion, which is then trapped by the nucleophile to form predominately the α -anomer, as depicted in Scheme 2. Following the glycosylation reactions, deacetylation products 6, 7, 11, and hydrogenation products 5, 6, 10, 12 were obtained in good yields under conventional conditions.

Most of these synthesized compounds were screened for their activity of inhibiting acute inflammatory response in the xylene-induced mouse ear edema model, and compared to HSS. ¹⁰ Intraperitoneal (ip) injection of compounds resulted in mice ear edema inhibition (Table 1). Ear weight reduction induced by compound **3b**, **7b**, and **10** were more significant than the control group.

Scheme 2. A proposed mechanism for non-Ferrier rearrangement reaction of 3,4,6-tri-O-acetyl-D-galactal.

Table 1. Effects of synthesized compounds on ear edema in mice^{a,b,c}

Compounds	Ear weight difference	Inhibition ratio (%)
	$mass \pm SD (mg)$	
Control	10.9 ± 5.7	
3a	12.0 ± 5.9	10.3
3b	$7.5 \pm 4.4^{*}$	49.6
4a	$8.6\pm2.1^*$	37.5
5b	$9.5 \pm 4.4^*$	36.1
6a	16.1 ± 6.0	0.0
7b	$6.1\pm2.8^*$	58.8
9	8.4 ± 1.8	13.0
10	$8.3 \pm 4.3^*$	45.2
11	9.9 ± 4.8	0.0
HSS	3.4 ± 1.4	69.0

^a Eight mice were used in each group.

The inhibition ratio on mice was 49.6%, 58.8%, and 45.2% respectively, with the dose of 100 mg/kg. The results suggested that these two 2-deoxy sugar derivatives exhibited considerable anti-inflammatory activity.

In conclusion, the synthesis of a series of *N*-phthalim-idomethyl 2,3-dideoxy- and 2,3-unsaturated sugar derivatives was accomplished. Three derivatives were found to have potent anti-inflammatory activity. It is interesting that the unsaturated and deoxy sugar functionalities in these molecules play quite different roles and this merits further investigation.

3. Experimental

3.1. General methods

Solvents were purified by standard procedures. Melting points were measured on an X4 melting point apparatus and are uncorrected. NMR spectra were recorded on Jeol-300 instrument (peaks marked with an asterisk indicate those exchangeable with D2O). Mass spectra were measured on an IBI-MDS Sciexciex Q-star or an FAB-MS mass spectrometer. Optical rotations were measured at 25 °C using an Optical Activity AA-10R automatic polarimeter. Elemental analyses were performed on a Perkin-Elmer 240 C instrument. TLC was performed on Silica Gel G_{F254} plates (Hai Yang Chemical Factory, Qingdao, Shandong, PR China) with diction by UV fluorescence quenching and by spraying with 10% H₂SO₄. Column chromatography was performed on Silica Gel H 60 (Hai Yang Chemical Factory, Qingdao, Shandong, PR China).

3.2. General procedure for compounds 3a, 3b, 4a, 9

To a stirred solution of glycal **2a**, **2b**, ⁶ or **8**¹¹ in dry toluene was added *N*-hydroxymethyl phthalimide **1**

(1.1 equiv) under argon at -20 °C, and then BF₃·Et₂O (1.5 equiv) was added dropwise. The reaction mixture was stirred at -20 °C for 1.5–6.0 h, and then neutralized with satd aq NaHCO₃. The organic layer was washed with water, dried over anhydrous Na₂SO₄, and evaporated. The residue was purified by column chromatography on silica gel to give the product.

3.3. General procedure for compounds 5a, 7a, 7b, 11

To a solution of compound **3a**, **3b**, **4a**, **10** in dry methanol was added sodium methoxide with stirring. The reaction mixture was stirred at room temperature for 0.5 h, the solvent was removed in vacuo, and the residue was purified by column chromatography on silica gel to give the product.

3.4. General procedure for compounds 5a, 5b, 6a, 6b, 10, 12

To a solution of compound 3a, 3b, 7a, 7b, 9, 11 in EtOAc was added 10% Pd/C (0.1 equiv) and the mixture was stirred under hydrogen at 1 atm for 8 h. The mixture was filtered and the filtrate concentrated in vacuo to give a residue that was purified by column chromatography on silica gel to give the product.

3.5. Phthalimidomethyl 4,6-di-*O*-acetyl-2,3-dideoxy-α-D-*threo*-hex-2-enopyranoside (3a)

Chromatographic purification (8:1, petroleum etheracetone) afforded 3a (60%) as a colorless solid; mp 101-102 °C; $[\alpha]_D - 161.9$ (c 0.4, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 2.07 (s, 6H, CH₃CO), 4.12 (dd, 2H, J = 6.9, 11.4 Hz H-6a), 4.26 (dd, 2H, J = 5.4, 11.4 Hz H-6b), 4.40 (m, 1H, H-5), 5.02 (m, 1H, H-4), 5.31 (m, 2H, CH₂), 5.41 (d, 1 H, H-1, $J_{1,2} = 2.7$ Hz), 5.98 (q, 1H, H-2, J = 3.3, 9.9 Hz), 6.13 (dd, 1 H, H-3, J = 5.4, 9.9 Hz), 7.80 (dd, 2H, J = 3.0, 5.7 Hz, Phth), 7.92 (dd, 2H, Phth); 13 C NMR (75 MHz, CDCl₃); δ 20.6, 20.7 (CH₃CO), 62.5 (C-6), 64.4 (C-5), 67.1 (C-4), 93.0 (C-1), 123.7 (C-3), 125.6 (C-2), 129.6, 131.8, 134.4 (Phth), 167.4 (C=O, Phth), 170.2, 170.6 (C=O, Ac); ESI-TOF-MS m/z: $[M+Na]^+$ 412.1; $[M+NH_4]^+$ 407.2. Anal. Calcd for C₁₉H₁₉NO₈ (389.1): C, 58.61; H, 4.92; N, 3.60. Found: C, 58.63; H, 4.94; N, 3.60.

3.6. Phthalimidomethyl 4,6-di-*O*-acetyl-2,3-dideoxy-α-D-*erythro*-hex-2-enopyranoside (3b)

Chromatographic purification (3:1, petroleum etheracetone) afforded **3b** (89%) as white needles; mp 120–121 °C; $[\alpha]_D$ +45.2 (c 1.2, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 2.08, 2.09 (2 s, 6H, 2Ac), 4.14–4.23 (m, 3H), 5.26–5.37 (m, 4H), 5.77–5.82 (m, 1H, H-2), 5.90 (d, 1H, H-3), 7.79 (dd, 2H, J = 3.0, 5.7 Hz,

^b Dosage 100 mg/kg.

 $^{^{\}rm c} p < 0.05$ Compared to control group by paired student's t-test.

Phth), 7.92 (dd, 2H, Phth); 13 C NMR (75 MHz, CDCl₃): δ 20.7, 20.9, 62.6, 64.7, 64.9, 67.2, 93.5, 123.7, 126.8, 129.7, 131.8, 134.5, 167.3, 170.2, 170.7; FAB-MS m/z: [M-H+K]⁺ 427.4. Anal. Calcd for C₁₉H₁₉NO₈ (389.1): C, 58.61; H, 4.92; N, 3.60. Found: C, 58.47; H, 4.74; N, 3.70.

3.7. Phthalimidomethyl 3,4,6-tri-*O*-acetyl-2-deoxy-α-D-*lyxo*-hexopyranoside (4a)

Colorless syrup; $[\alpha]_D + 0.12$ (c 0.9, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 1.96, 2.04, 2.09 (3s, 9H, 3 × Ac), 2.14–2.19 (m, 2H, H-2), 4.00 (dd, 1H, J = 5.4, 11.1 Hz, H-6a), 4.12 (dd, 1H, J = 3.6, 11.1 Hz, H-6b), 4.26 (t, 1H, J = 6.6 Hz, H-5), 5.19–5.34 (m, 4H), 7.79 (dd, 2H, J = 2.7, 5.7 Hz, Phth), 7.93 (dd, 2H, Phth); ¹³C NMR (75 MHz, CDCl₃): δ 20.6, 20.6, 20.7 (CH₃CO), 29.7 (C-6), 62.0 (O–CH₂–N), 63.9 (C-5), 65.7 (C-4), 66.4 (C-3), 67.1 (C-2), 96.5 (C-1), 123.8, 131.7, 134.5 (Phth), 167.3 (C=O, Phth), 169.8, 170.2, 170.4 (C=O, Ac); ES1-TOF-MS m/z: $[M+NH_4]^+$ 467.2, $[M+Na]^+$ 472.1. Anal. Calcd for $C_{21}H_{23}NO_{10}$ (449.1): C, 56.12; H, 5.16; N, 3.12. Found: C, 56.33; H, 4.99; N, 3.10.

3.8. Phthalimidomethyl 4,6-di-*O*-acetyl-2,3-dideoxy-α-D-*threo*-hexopyranoside (5a)

Chromatographic purification (3:1, petroleum etheracetone) afforded **5a** (75%) as a white foam; $[\alpha]_D + 79.9$ (c 2.0, CHCl₃); 1 H NMR (300 MHz, CDCl₃): δ 1.55–2.00 (m, 4H, H-2, H-3), 2.03 (s, 3H, CH₃CO), 2.08 (s, 3H, CH₃CO), 3.97 (dd, 1H, J = 6.9, 11.1 Hz, H-6a), 4.10 (dd, 1H, J = 6.0, 11.1 Hz, H-6b), 4.21 (m, 1H, H-5), 4.92 (s, 1H, H-1), 5.19–5.29 (m, 4H), 7.80 (dd, 2H, Phth), 7.92 (dd, 2H, Phth); 13 C NMR (75 MHz, CDCl₃): δ 20.7, 21.2, 22.0, 23.8, 61.5 (O–CH₂–N), 63.2, 63.9, 66.4, 67.5, 95.9 (C-1), 123.7, 131.8, 134.4 (Phth), 167.6 (C=O, Phth), 170.4, 170.7 (C=O, Ac); ESI-TOF-MS m/z: [M+Na]⁺ 413.1. Anal. Calcd for $C_{19}H_{21}NO_8$ (391.1): C, 58.31; H, 5.41; N, 3.58. Found: C, 58.13; H, 5.24; N, 3.40.

3.9. Phthalimidomethyl 4,6-di-*O*-acetyl-2,3-dideoxy-α-D-*erythro*-hexopyranoside (5b)

Chromatographic purification (3:1, petroleum etheracetone) afforded **5b** (75%) as a white foam; [α]_D +53.3 (c 1.3, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 1.79–2.00 (m, 4H, H-2, H-3), 2.04 (s, 3H, CH₃CO), 2.08 (s, 3H, CH₃CO), 4.01–4.09 (m, 2H), 4.22–4.30 (m, 1H), 4.70–4.75 (m, 1H), 5.15 (s, 1H, H-1), 5.22 (d, 1H, J = 10.2 Hz, CH₂), 5.28 (d, 1H, CH₂), 7.80 (dd, 2H, Phth), 7.92 (dd, 2H, Phth); ¹³C NMR (75 MHz, CDCl₃): δ 20.7, 21.2, 22.0, 23.8, 61.5 (O–CH₂–N), 62.8, 63.9, 67.4, 69.1, 95.6 (C-1), 123.7, 131.8, 134.4 (Phth), 167.5 (C=O, Phth), 170.0, 170.9 (C=O, Ac); ESI-TOF-MS

m/z: [M+NH₄]⁺ 409.2. Anal. Calcd for C₁₉H₁₉NO₈ (391.1): C, 58.31; H, 5.41; N, 3.58. Found: C, 58.33; H, 5.54; N, 3.32.

3.10. Phthalimidomethyl 2,3-dideoxy-α-D-threo-hexo-pyranoside (6a)

Chromatographic purification (1:1, petroleum etheracetone) afforded **6a** (82%) as a white powder; mp 117–119 °C; $[\alpha]_D$ –68.2 (c 1.1, CH₃OH); ¹H NMR (300 MHz, DMSO- d_6): δ 3.34–3.37 (m, 1H, H-6a), 3.49–3.52 (m, 1H, H-6b), 3.55–3.61 (m, 1H, H-5), 3.67–3.72 (m, 1H, H-4), 4.09 (t, 1H, OH)*, 4.54 (d, 1H, OH)*, 5.12 (d, 1H, J=11.4 Hz, H-1), 5.20 (m, 2H, CH₂), 5.71 (q, 1H, H-2), 5.92 (q, 1H, H-3), 7.91–7.99 (m, 4H, Phth); ¹³C NMR (125 MHz, DMSO- d_6): δ 60.7 (O–CH₂–N), 60.9 (C-6), 64.3 (C-5), 72.5 (C-4), 94.2 (C-1), 24.2, 27.3 (C-2, C-3), 130.7, 132.2, 135.1 (Phth), 168.9 (C=O); ESI-TOF-MS m/z: [M+NH₄]* 325.1; [M+Na]* 330.1. Anal. Calcd for C₁₅H₁₇NO₆ (307.1): C, 58.63; H, 5.58; N, 4.56. Found: C, 58.72; H, 5.52; N, 4.53.

3.11. Phthalimidomethyl 2,3-dideoxy-α-D-*erythro*-hexopyranoside (6b)

[α]_D +87.5 (c 1.0, CH₃OH). White foam; ¹H NMR (300 MHz, D₂O): δ 1.44–1.73 (m, 4H, H-2, H-3), 3.34–3.40 (m, 4H), 4.93 (s, 1H), 5.02 (d, 1H, J = 11.7 Hz, O–CH₂–N), 5.07 (d, 1H, O–CH₂–N), 7.61–7.64 (m, 4H, Phth); ¹³C NMR (75 MHz, D₂O): δ 26.4, 28.8, 61.2, 64.9, 65.5, 74.8, 96.7 (C-1), 124.3, 131.5, 135.7, 170.0 (C=O); ESI-TOF-MS m/z: [M+H]⁺ 308.1; [M+Na]⁺ 330.1. Anal. Calcd for C₁₅H₁₇NO₆ (307.1): C, 58.63; H, 5.58; N, 4.56. Found: C, 58.42; H, 5.35; N, 4.83.

3.12. Phthalimidomethyl 2,3-dideoxy-α-D-threo-hex-2-enopyranoside (7a)

Chromatographic purification (1:1, petroleum etheracetone) afforded 7a (82%) as a white powder; mp 123-125 °C; $[\alpha]_D -55.3$ (c 1.0, CH₃OH); ¹H NMR (300 MHz, DMSO- d_6): δ 3.24–3.33 (m, 1H, H-6a), 3.45-3.62 (m, 1H, H-6b), 3.64-3.67 (m, 1H, H-5), 3.70-3.74 (m, 1H, H-4), 4.42 (t, 1H, J = 5.4 Hz, OH)*, 4.67 (d, 1H, J = 7.8 Hz, OH)*, 5.06 (d, 1H, H-1, $J = 11.4 \text{ Hz}, \text{ CH}_2$, 5.18 (s, 1H, H-1), 5.19 (m, 1H, CH_2), 5.73 (dd, 1H, J = 3.0, 10.2 Hz, H-2), 5.99 (dd, 1H, J = 5.4, 10.2 Hz, H-3), 7.86–7.94 (m, 4H, Phth); ¹³C NMR (75 MHz, DMSO- d_6): δ 60.2 (O–CH₂–N), 60.9 (C-6), 65.0 (C-5), 72.3 (C-4), 94.0 (C-1), 124.2, 127.4, 130.9, 132.0, 135.7 (Phth), 168.4 (C=O); ESI-TOF-MS m/z: $[M+NH_4]^+$ 323.2; $[M+Na]^+$ 328.1. Anal. Calcd for C₁₅H₁₅NO₆ (305.1): C, 59.01; H, 4.95; N, 4.59. Found: C, 58.79; H, 5.07; N, 4.30.

3.13. Phthalimidomethyl 2,3-dideoxy-α-D-*erythro*-hex-2-enopyranoside (7b)

White foam, $[\alpha]_D + 58.6$ (c 0.7, CH_3OH); 1H NMR (300 MHz, DMSO- d_6): δ 3.44–3.51 (m, 3H, H-5, H-6), 3.90 (t, 1H, OH)*, 4.50 (t, 1H, OH)*, 5.05 (d, 1H, J = 6.9 Hz), 5.10 (d, 1H, J = 11.1 Hz, CH_2), 5.17 (s, 1H), 5.19 (d, 1H, CH_2), 5.59–564 (m, 1H, H-2), 5.85 (d, 1H, H-3), 7.87–7.95 (m, 4H, Phth); ^{13}C NMR (75 MHz, DMSO- d_6): δ 60.5 (O– CH_2 –N), 61.9 (C-6), 64.3 (C-5), 73.0 (C-4), 92.9 (C-1), 123.5, 124.7, 131.5, 134.8, 135.0 (Phth), 167.4 (C=O); ESI-TOF-MS m/z: [M+NH₄]* 323.1. Anal. Calcd for $C_{15}H_{15}NO_6$ (305.1): C, 59.01; H, 4.95; N, 4.59. Found: C, 58.72; H, 5.12; N, 4.53.

3.14. Phthalimidomethyl 2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl- $(1\rightarrow 4)$ -6-O-acetyl-2,3-dideoxy- α -D-erythro-hex-2-enopyranoside (9)

Chromatographic purification (1:1, petroleum etheracetone) afforded 9 (71%) as a white solid; mp 92-93 °C; $[\alpha]_D + 31.7$ (c 1.3, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 1.97, 2.05, 2.07, 2.10, 2.15 (5s, 15H, CH₃CO), 3.92 (t, 1H, J = 6.6 Hz), 4.06-4.28 (m, 6H), 4.56 (d, 1H, H-1', J = 7.8 Hz), 5.00 (dd, 1H, J = 3.6, 11.5 Hz), 5.19 (dd, 1H, J = 7.8, 10.5 Hz), 5.24 (d, 1H, J = 10.2 Hz, CH₂), 5.30 (d, 1H, CH₂), 5.30–5.32 (m, 1H), 5.37 (d, 1H, J = 2.7 Hz, H-1), 5.69–5.73 (m, 1H, H-2), 6.10 (d, 1H, J = 10.2 Hz, H-3), 7.78 (dd, J = 3.0, 5.4 Hz, 2H, Phth), 7.91 (dd, 2H, Phth); ¹³C NMR (75 MHz, CDCl₃): δ 20.4, 20.5, 20.7 (CH₃CO), 61.1, 62.7, 64.6, 66.7, 67.7, 68.6, 70.6, 70.7, 72.8, 93.4 (C-1), 102.0 (C'-1), 123.7, 126.0 (C-2, C-3), 131.7, 132.0, 134.4 (Phth), 167.4 (C=O, Phth), 167.4, 169.3, 170.0, 170.2, 170.6 (C=O, Ac); FAB-MS m/z: $[M+K]^+$ 716.3. Anal. Calcd for C₃₁H₃₅NO₁₆ (677.2): C, 54.95; H, 5.21; N, 2.07. Found: C, 54.80; H, 5.33; N, 2.00.

3.15. Phthalimidomethyl 2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl- $(1\rightarrow 4)$ -6-O-acetyl-2,3-dideoxy- α -D-erythro-hexopyranoside (10)

Chromatographic purification (5:2, petroleum etheracetone) afforded **10** (77%) as a colorless syrup; $[\alpha]_D + 36.2$ (c 1.7, CHCl₃); 1 H NMR (300 MHz, CDCl₃): δ 1.74–1.86 (m, 4H), 1.98, 2.03, 2.07, 2.10, 2.15 (5s, 15H, CH₃CO), 3.53–3.58 (m, 1H), 3.89–3.98 (m, 2H), 4.05–4.23 (m, 4H), 4.53 (d, 1H, J = 8.1 Hz, H-1′), 4.98 (dd, 1H, J = 3.0, 10.5 Hz), 5.09–5.27 (m, 4H), 5.37 (d, 1H, J = 3.3 Hz), 7.78 (dd, 2H, J = 3.0, 5.7 Hz, Phth), 7.92 (dd, 2H, Phth); 13 C NMR (75 MHz, CDCl₃): δ 20.4, 20.5, 20.6, 20.8 (5CH₃CO), 25.7, 28.5, 61.1, 63.2, 63.7, 66.7, 68.6, 69.4, 70.4, 70.8, 76.1, 95.3 (C-1), 102.0 (C′-1), 123.7, 131.7, 134.4 (Phth), 167.4, 169.4 170.0, 170.2, 170.3, 170.7 (C=O); ESI-TOF-MS m/z: [M+NH₄] $^+$ 697.2, [M+Na] $^+$ 702.2. Anal. Calcd for

C₃₁H₃₇NO₁₆ (679.2): C, 54.79; H, 5.49; N, 2.06. Found: C, 54.70; H, 5.47; N, 1.99.

3.16. Phthalimidomethyl β -D-galactopyranosyl- $(1\rightarrow 4)$ -2,3-dideoxy- α -D-*erythro*-hex-2-enopyranoside (11)

Chromatographic purification (15:1, CHCl₃–MeOH) afforded **11** (80%) as a colorless syrup; $[\alpha]_D + 62.7$ (c 1.0, CH₃OH); ¹H NMR (300 MHz, DMSO- d_6): δ 3.25–3.56 (m, 9H), 4.09–4.19 (m, 2H, H-1, H'-1), 4.40 (d, 1H, OH)*, 4.53–4.57 (m, 2H, $2 \times OH$)*, 4.74 (s, 1H, OH)*, 4.96 (s, 1H, OH)*, 5.10–5.21 (m, 3H, O–CH₂–N, H'-3), 5.64 (d, 1H, J = 10.2 Hz, H-2), 6.05 (d, 1H, J = 10.2 Hz, H-3), 7.87–7.95 (m, 4H, Phth); ¹³C NMR (75 MHz, DMSO- d_6): δ 60.0, 60.4, 64.4, 68.1, 70.6, 71.2, 73.2, 75.2, 79.2, 93.0 (C-1), 105.0 (C'-1), 123.5, 125.4, 131.5, 133.4, 134.8, 167.4 (C=O); ES1-MS-MS m/z: $[M+Na]^+$ 490.1, $[M+NH_4]^+$ 485.2, $[M+H]^+$ 468.2. Anal. Calcd for $C_{21}H_{25}NO_{11}$ (467.1): C, 53.96; H, 5.35; N, 3.00. Found: C, 53.63; H, 5.49; N, 2.96.

3.17. Phthalimidomethyl β -D-galactopyranosyl- $(1\rightarrow 4)$ -2,3-dideoxy- α -D-*erythro*-hexopyranoside (12)

Chromatographic purification (15:1, CHCl₃–MeOH) afforded **12** (71%) as a colorless syrup; $[\alpha]_D$ +28.3 (c 0.8, CH₃OH); ¹H NMR (300 MHz, D₂O): δ 1.59–1.68 (m, 3H), 1.90–1.93 (m, 1H), 3.26–3.32 (m, 1H), 3.39–3.58 (m, 8H), 3.73 (d, 1H, J = 3.0 Hz), 4.26 (d, 1H, J = 7.5 Hz, H'-1), 4.96 (s, 1H), 5.08 (d, 1H, J = 11.4 Hz, O–CH₂–N), 5.11 (d, 1H, O–CH₂–N), 7.67–7.75 (m, 4H, Phth); ¹³C NMR (75 MHz, D₂O): δ 25.7, 28.4, 50.1, 62.1, 62.8, 65.7, 66.6, 68.2, 69.6, 70.5, 77.7, 92.0 (C-1), 102.1 (C'-1), 121.4, 126.8, 130.0 (Phth), 169.2 (C=O, Phth); ESI-TOF-MS m/z: $[M+NH_4]^+$ 487.2, $[M+Na]^+$ 482.2. Anal. Calcd for C₂₁H₂₇NO₁₁ (469.2): C, 53.73; H, 5.80; N, 2.98. Found: C, 53.83; H, 6.00; N, 3.11.

3.18. General methods for determination of anti-inflammation activity. 10

Experimental animals were ICR (Institute of Cancer Research) mice provided by the Center for Experimental Animals, Peking University Health Science Center. Only male mice (SPF grade, 4–6 weeks old and weighing between 18 and 22 g) were used in the study. Mice were allowed free access to food and drink and housed at room temperature and at a humidity of 45–55%. Hydrocortisone sodium succinate (HSS) was purchased from Tianjin Biochemical Pharmaceuticals.

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References

- Chan, C. L.; Lien, E. J.; Tokes, Z. A. J. Med. Chem. 1987, 30, 509-514.
- Bailleux, V.; Vallee, L.; Nuyts, J. P.; Vamecq, J. Biomed. Pharmacother. 1994, 48, 95–101.
- Lima, L. M.; Castro, P.; Machado, A. L.; Fraga, C. A. M.; Lugnier, C.; Concalves de Moraes, V. L.; Barreiro, E. J. Bioorg. Med. Chem. 2002, 10, 3067–3073.
- 4. Hashimoto, Y. Bioorg. Med. Chem. 2002, 10, 461-479.
- Srivastava, R. M.; Oliveira, F. J. S.; Da Silva, L. P.; De Freitas Filho, J. R.; Oliveira, S. P.; Lima, V. L. M. Carbohydr. Res. 2001, 332, 335–340.

- Danishefsky, S. J.; Bilodeau, M. T. Angew. Chem., Intl. Ed. 1996, 35, 1380–1419.
- Ferrier, R. J.; Overend, W. G.; Ryan, Miss A. E. J. Chem. Soc. 1962, 3667–3670.
- 8. Lin, H.; Du, W.; Chang, C.; Lin, C. Tetrahedron Lett. **2005**, 46, 5071–5076.
- 9. Wallenfels, K.; Lehmann, J. Ann. 1960, 635, 166-177.
- Shen, P. N.; Ruan, K. F.; Wang, Yu L.; Yu, W.; Zhang, W. Q.; Hong, X. K.; Wang, X. H. U.S. Patent 6,787,165, 2004.
- Stick, R. V.; Stubbs, Keith A.; Tilbrook, D.; Matthew, G.; Watts, A. G. Aust. J. Chem. 2002, 83–86.